

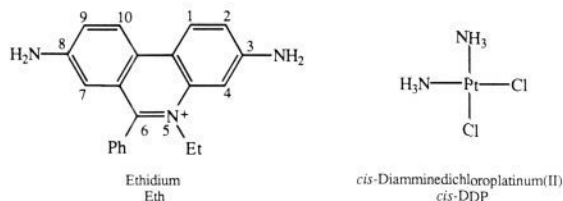
## DNA Promotes the Reaction of *cis*-Diamminedichloroplatinum(II) with the Exocyclic Amino Groups of Ethidium Bromide

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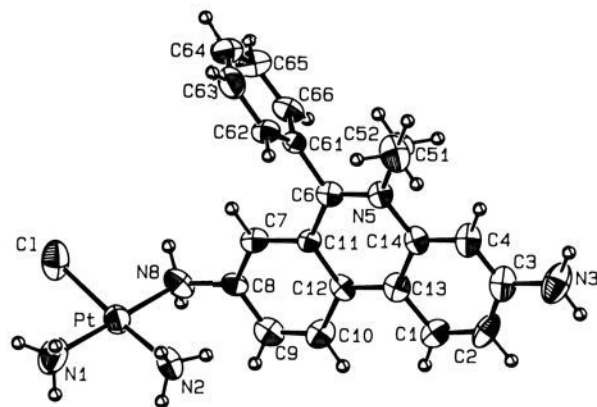
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DNA functions as a template for replication and transcription of genetic information in cells. Duplex DNA can also provide a one-dimensional surface that modulates the reaction of small molecules<sup>1</sup> or directs the diffusion of macromolecules.<sup>2</sup> Here we report details of a chemical reaction between the antitumor drug *cis*-diamminedichloroplatinum(II) (*cis*-DDP) and the ethidium

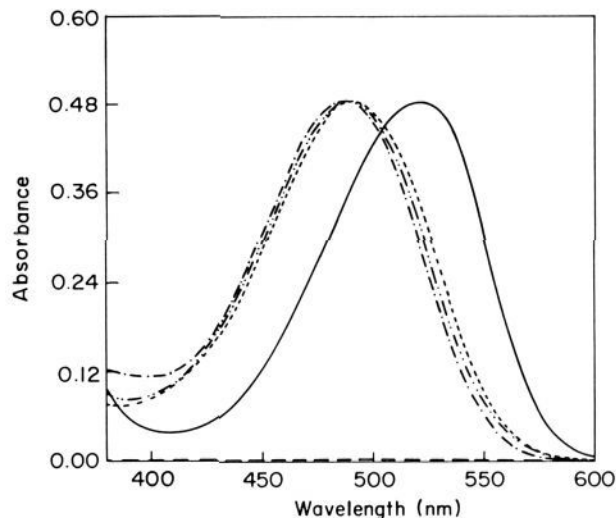


cation (Eth) that is promoted by DNA. In this chemistry, a *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>Cl]<sup>+</sup> unit coordinated at a purine N7 site on DNA is ideally positioned to react with an ethidium molecule intercalated between the adjacent base pairs. The result is a metastable complex in which the *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>]<sup>2+</sup> moiety covalently links the 3- or 8-amino group of Eth and the DNA nucleobase.

Duplex DNA dramatically increases the extent of reaction between *cis*-DDP and ethidium bromide.<sup>3</sup> The nature of the Pt–Eth–DNA ternary complex so formed has been uncertain, however, since the complex is rather unstable, slowly losing ethidium upon incubation at 37 °C (*t*<sub>1/2</sub> ≈ 10 h). The recent synthesis and structural characterization of *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N9-9-AA)Cl](NO<sub>3</sub>),<sup>4</sup> in which platinum is coordinated to the exocyclic amino group of 9-aminoacridine (9-AA), suggested that platinum might also bind to the exocyclic amino groups of ethidium to form complexes of the general formula *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(Eth)Cl]<sup>2+</sup> that could be characterized chemically and subsequently bound to DNA. As anticipated, these platinum–ethidium precursor complexes form in the reaction of ethidium nitrate and [Pt(NH<sub>3</sub>)<sub>2</sub>Cl]<sup>+</sup>, L = NO<sub>3</sub><sup>−</sup> (*n* = 0) or DMF (*n* = 1+), in DMF.<sup>5</sup> This reaction produces, after HPLC purification, the acetate salts *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N3-Eth)Cl](OAc)<sub>2</sub> (**1**) and *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N8-Eth)Cl](OAc)<sub>2</sub> (**2**).<sup>6</sup> The identity of the purified complexes has been unambiguously determined by <sup>1</sup>H NMR, <sup>195</sup>Pt NMR, and mass spectrometric studies.<sup>6</sup> The structure of the N8 linkage isomer has also been determined by X-ray diffraction (Figure 1).<sup>6,7</sup>



**Figure 1.** Structure of *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N8-Eth)Cl]<sup>2+</sup> showing the 40% thermal ellipsoids (except for hydrogen) and atom labeling scheme. Selected bond distances (Å) and angles (deg) are as follows: Pt–Cl, 2.280 (9); Pt–N1, 2.03 (2); Pt–N2, 2.11 (2); Pt–N8, 2.05 (1); N3–C3, 1.39 (2); N8–C8, 1.46 (2); Pt–N8–C8, 118.0 (8). Only one of the two disordered positions for the platinum coordination plane is depicted (see Supplementary Material).



**Figure 2.** Optical absorption spectra of ethidium–DNA complexes: (—) ethidium + DNA before *n*-butanol extraction, (---) ethidium + DNA after extraction, (- - -) *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N3-Eth)Cl](OAc)<sub>2</sub> (**1**) + DNA after extraction, (· · ·) *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N8-Eth)Cl](OAc)<sub>2</sub> (**2**) + DNA after extraction, (- · - ·) ethidium + *cis*-DDP + DNA after extraction. Experimental details are given in Supplementary Material.

The optical absorption spectra of the DNA adducts of **1** and **2** are shown in Figure 2. These spectra are quite distinct from that of intercalatively bound ethidium but almost exactly match the absorption spectrum of the ternary complex formed by *cis*-DDP and ethidium on DNA (Figure 2).<sup>6</sup> Furthermore, ethidium is slowly released from DNA complexes of **1** and **2** upon incubation at 37 °C. These data demonstrate that the precursor ethidium–platinum complexes **1** and **2** form the same ternary complexes on DNA as obtained in the reaction of *cis*-DDP with ethidium bromide in the presence of DNA. The ternary complexes therefore must consist of platinum coordinated by two *cis* amines, an N3- or N8-bound ethidium, and a DNA ligand, presumably the N7 atom of either guanine or adenine.<sup>8</sup>

We assume that the Pt–Eth–DNA complex forms via ethidium intercalation, known to be rapid,<sup>9</sup> followed by monofunctional binding of *cis*-DDP to DNA. To model this intermediate, a molecular mechanics energy minimized structure<sup>10</sup> of the duplex DNA hexamer, d[C(1)pG(2)pC(3)pG(4)pC(5)pG(6)]-d[C(7)-

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(7) Crystal data for *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N8-Eth)Cl](ClO<sub>4</sub>)<sub>2</sub>·MeOH·0.5H<sub>2</sub>O, C<sub>22</sub>H<sub>31</sub>N<sub>5</sub>O<sub>9</sub>Cl<sub>3</sub>Pt, *M*<sub>r</sub> = 818.97, triclinic, space group *P*1̄, *a* = 13.269 (7) Å, *b* = 16.125 (3) Å, *c* = 8.156 (5) Å, α = 99.93 (4)°, β = 96.29 (6)°, γ = 112.29 (3)°, *V* = 1561 (3) Å<sup>3</sup>, *Z* = 2, ρ<sub>calcd</sub> = 1.741 gm/cm<sup>3</sup>. For 2697 observed reflections and 376 variables, the current discrepancy indices are *R* = 0.052, *R*<sub>w</sub> = 0.068.

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**Figure 3.** Proposed intermediate in the DNA-promoted reaction of *cis*-DDP with ethidium. This model is comprised of a duplex DNA hexamer, d[C(1)pG(2)pC(3)pG(4)pC(5)pG(6)]-d[C(7)pG(8)pC(9)pG(10)pC(11)pG(12)], in which ethidium is intercalated between the C(3)-G(10) and G(4)-C(9) base pairs, and an idealized *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>Cl]<sup>+</sup> fragment is coordinated to N7 of G(4) (see text for discussion). The two reacting groups, -NH<sub>2</sub> and Pt, are highlighted in yellow.

pG(8)pC(9)pG(10)pC(11)pG(12)], in which ethidium is intercalated between the C(3)-G(10) and G(4)-C(9) base pairs,<sup>11</sup> was linked to an idealized *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>Cl]<sup>+</sup> fragment at N7 of G(4).<sup>12</sup> The resulting structure (Figure 3) illustrates how DNA could serve as a template to orient the ethidium N8 amino group above the square planar platinum complex (Pt—N8-Eth = 3.9 Å, N7-G(4)—Pt—N8-Eth = 62°) in the major groove of the double helix to effect the nucleophilic displacement of the chloride ligand. A similar model in which platinum is coordinated to N7 of G(10) on the other DNA strand reveals a favorable orientation for reaction with the N3 amino group of ethidium, although the distance is somewhat greater (Pt—N3-Eth = 4.7 Å).

DNA does not promote the analogous reaction of *trans*-DDP with ethidium.<sup>3</sup> Model studies reveal that it is stereochemically impossible for the exocyclic amino groups of intercalated ethidium to bind *trans* to a coordinated nucleobase in duplex DNA. The reaction of *cis*-DDP with proflavin, however, is facilitated by DNA.<sup>3</sup> A model based upon the crystal structure of proflavin intercalated into the duplex dimer (CpG)<sub>2</sub><sup>13</sup> reveals the exocyclic amino groups of proflavin, like those of ethidium, to be in a favorable position to react with platinum bound to N7 of guanine in the major groove of DNA. Acridine,<sup>3</sup> which has no exocyclic amino group, and 9-aminoacridine,<sup>14</sup> which intercalates with the N9 amino group projecting into the minor groove,<sup>15</sup> do not react with *cis*-DDP in the presence of DNA. The model in which the *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>Cl]<sup>+</sup> unit is bound to N7 of a purine base in the major

groove of DNA at a site adjacent to the exocyclic amino group of the intercalator explains all of these observations.

In the absence of DNA, 3.2 mM *cis*-DDP and ethidium bromide react to consume 21% of the intercalator in 8 h; at 50 μM concentrations, less than 2% of the ethidium reacts after 8 or 24 h.<sup>6</sup> In the presence of DNA, however, 19.9% of the ethidium reacts in 8 h at 50 μM concentrations of *cis*-DDP and ethidium bromide (*r<sub>f</sub>* 0.25).<sup>6</sup> The reaction of *cis*-DDP with EthBr therefore proceeds to the same extent at 60-fold lower concentrations of both reactants when carried out in the presence of DNA.

In conclusion, the present results reveal details of how DNA can promote the reaction of a Lewis acid (platinum) with a Lewis base (Eth exocyclic amino group) under conditions where no significant reaction would occur in its absence. This acid-base chemistry<sup>3</sup> joins electron transfer and macromolecular diffusion as a general class of reactions facilitated by DNA.

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**Supplementary Material Available:** Experimental synthetic, mechanistic, spectroscopic, analytical, and reaction chemistry details as well as tables of atomic positional and thermal parameters for *cis*-[Pt(NH<sub>3</sub>)<sub>2</sub>(N8-Eth)Cl](ClO<sub>4</sub>)<sub>2</sub>·MeOH·0.5H<sub>2</sub>O (5 pages). Ordering information is given on any current masthead page.

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